

# B4

## An Organ Slice Model to Evaluate Carcinoid Tumorigenesis in Liver

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**Background:** Midgut carcinoid cancers typically metastasize to the liver. However, cancer cell interactions with the liver tissue microenvironment are poorly understood. In combination with live-cell imaging methods and multi-photon microscopy, we developed an organ slice culture system that more closely resembles the three-dimensional, multicellular tumor microenvironment than does a dispersed cell culture system. Using this system we assessed cancer cell dynamics *in situ* and demonstrated that tumor-like structures are formed as a result of two concurrent processes — cell proliferation and aggregation.

**Methods:** Human carcinoid cancer cells, stably transfected with GFP were introduced to intact mouse liver via hepatic portal vein injection. A long-lasting vital dye was also introduced to mark the vasculature. The liver was sectioned into 200  $\mu\text{m}$  thick slices and maintained in culture for up to two weeks. Cancer cell dynamics were assessed by immunofluorescence methods or by time-lapse imaging.

**Results:** The formation of tumor-like structures was observed by the 4<sup>th</sup> to 6<sup>th</sup> day in culture and it was revealed that more than half of the carcinoid cells were still dividing in the liver slices. Moreover, we found that tumor cells exhibited a variety of characteristic movements and rapidly inter-converted between elongated and rounded (blebbing) modes of locomotion. A rapid form of ameoboid movement, observed only after several days in culture, was correlated to tumor cell aggregation in slices. These observations were consistent with *in vitro* experiments that demonstrated that liver tissue pieces or conditioned medium promoted carcinoid cancer cell aggregation in culture in a dose-dependent manner.

**Conclusion:** Proliferation and aggregation induced by an unidentified tissue-specific soluble factor contributed to the

formation of tumor-like structures in mouse liver slices. This *ex vivo* model provides a tractable and cost efficient system for studying tumor formation in mouse liver.